

# **1. Phosphorus Concentrations in Environmental Samples**

# 1.8 Plant and Animal Biomass

Karen Baumann, Dana Zimmer, Rhena Schumann

## 1.8.1 Plants

Depending on their habitat the mean TP concentrations for terrestrial plants are 2 g kg<sup>-1</sup> and for aquatic plants 6 g kg<sup>-1</sup> (Lerch 1990) (Tab. 1.8-1). This can be attributed to the higher dry matter of terrestrial plants (more supporting tissue from cellulose and lignin) compared to aquatic plants (Duchesne & Larson 1989). Concentrations are also affected by external factors such as P availability in soil. Under favourable conditions, sunflowers accumulated 2.5 times more P in their leaves (up to 7.2 mmol TP m<sup>-2</sup>) in comparison to half of the P availability (2.0 mmol TP m<sup>-2</sup>, Jacob & Lowlor 1991). TP concentrations also depend on plant specific factors (e. g. plant species, genotype) and are organ-specific within the plant. For example, TP concentration in fine roots of copper beech was twice as high as in wood, bark and twigs (Lerch 1990). Plant age and season affect TP concentrations as well, since in different development stages different amounts of P are necessary (Wang et al. 2015, Saunders & Metson 1971).

Species or group	Tissue	influencing factors	TP (g kg <sup>-1</sup> DM <sup>-1</sup> )	Reference
Terrestrial plants		Habitat, degree of organisation (supporting tissue)	0.5-8	Lerch (1991)
Aquatic plants			6	Lerch (1991)
<b>Spruce</b> (Pinus sylvestris)	Leaves	Species specific, normal nutrient	1.3-1.9	Mellert & Göttlein
Fir ( <i>Picea abies</i> )		supply	1.5-2.2	(2012)
Beech (Fagus sylvatica)		,	1.2-1.9	· · ·
<b>Oak</b> (Quercus spp.)			1.4-2.1	
<b>Copper beech</b> (F. sylvatica)	Fine roots	Different tissues	2.2	Lerch (1991)
	Wood Bark Twigs		0.9	
			0.9	
			0.9	
	leaves		1.3	

#### **Table 1.8-1** TP Concentrations in plant tissues and macro-algae (DM = dry matter)

Handbook on the selection of methods for digestion and determination of total P in environmental samples





## 1.8.2 Animals

Similar to plants, the TP concentrations in animals depend on nutrition, tissue, ages and development stage. For a lot of vertebrates, and hence also for farm animals, especially the Ca:P ratio is important. For this reason, lots of studies demonstrate that P availability and feed utilization are affected by Ca concentration in feed (e.g. Song et al. 2017). On the one hand, P absorption can be inhibited by Ca, if slightly soluble Ca-phosphates are in the diet, which cannot be dissolved and adsorbed (Nakamura 1982). On the other hand, high P concentrations in feed can reduce Ca accumulation in bones (Masuyama et al. 2003). Song et al. (2017) demonstrated that activity of phosphatases in blood serum is a crucial factor for mineralisation of bones and accumulation of Ca and P in fish scales. In fishes the fish scales can be a sink for Ca and P, supporting Ca and P homoeostasis (Song et al. 2017, see table 1.8-3).

The way of life (herbivore/carnivore) and the feed availability depending on season (and therefore food sources) have an important impact on TP concentrations in animal bodies (Ghaddar & Saoud 2012). These authors documented highest TP concentration in muscular meat of white seabream in April but lowest in June (see table 1.8-2). This interrelation could be used to use special animals in special seasons for P poor diet of humans (e.g. some kidney diseases). TP concentrations vary especially with animal species (table 1.8-2) but also with tissues (blood, serum, muscles, bones).

Species or	Tissue	Influen-	TP in	TP in	Reference
group	IISSUE	cing	tissues	serum	Reference
		factors	(g kg <sup>-1</sup> DM <sup>-1</sup> )	(g l <sup>-1</sup> )	
Coral (Lophelia	body	recent	0.016		Mason et al.
pertusa)	,	fossil	0.123		(2011)
Rabbit fish (Siganus rivulatus)	flesh	herbivore	8.95		Ghaddar & Saoud
White sea-		carnivore	11.32		(2012)
<b>bream</b> (Diplodus sargus)					
Pheasant	Pectoral		10.16		Straková et
(Phasianus colchicus)	muscle	specific			al. (2011)
Broiler	Pectoral		9.25		
	muscle				
Cat	Muscle		10.5		Cuthbertson (1925)
Pig	Muscle		6.03		Jastrzębska et al. (2010)
Sheep Merino	Muscle,	Mass:			Bellof et al.
Landsheep	ages	18 kg	8.09		(2006)
		55 kg	6.17		
Human	Muscle		1.56		Forbes et al. (1953)
Japanese seabass (La-	Serum	31 g Ca kg feed <sup>-1</sup>		0.43	Song et al. (2012)
teolabrax japonicus)		4.2 g Ca kg feed <sup>-1</sup>		0.31	
Human	Blood			0.36 - 0.43	Kay & Byrom (1927)

Table 1.8-2 TP (	concentrations in	animal	tissues or serum	(DM = dry matter)	

In humans high TP concentrations can be found in teeth and bones. According to Koolmann & Röhm (1998), the mean TP concentration in humans is 10 g kg<sup>-1</sup> body mass and the daily TP demand is 0.8 g d<sup>-1</sup>. Whereas in vertebrates high TP concentrations are in the endoskeleton, in invertebrates, such as mussels, gastropods and corals, most P is concentrated in the exoskeleton (table 1.8-3). For animals the growth stage is relevant for TP concentrations, for example lambs (German Merino Landsheep) have highest TP concentrations in early development stages, which decreases with development (Bellof et al. 2006). This decrease in TP concentrations can be attributed to the decrease in water concentrations in bone tissue during growth, which increases dry matter concentration in bones of older animals (Bellof et al. 2006).

Species or group	Suppor- ting tissue	Influen- cing factors	TP (g kg <sup>-1</sup> DM <sup>-1</sup> )	Reference
<b>Oyster</b> (Ostreidae)	Shell		0.9	Yoon et al. (2003)
<b>Snails</b> (Archacha- tina, Achatina spp.)	Shell		10-69	Ademolu et al. (2016)
Japanese	Vertebrae	31 g Ca kg	125	Song et al.
seabass (La-	Scale	Futter <sup>-1</sup>	74	(2012)
teolabrax japonicus)	Vertebrae	4,2 g Ca kg	138	
	Scale	Futter <sup>-1</sup>	87	
Human	Teeth		125-137	Hennequin et al. (1994)
	dry, non- lipid bones		9.2-10	Zipkin et al. (1960)
	Bone ash		171-175	

**Table 1.8-3** TP concentrations in animal skeletons (DM = dry matter)

#### References

- Ademolu K, Precious O, Ebenso I, Babatunde I (2016) Morphometrics and mineral composition of shell whorls in three species of giant Africans snails from Abeokuta, Nigeria. Folia Malacol 24: 81-84, DOI: <u>10.12657/folmal.024.013</u>
- Bellof G, Most E, Pallauf J (2006) Concentration of Ca, P, Mg, Na and K in muscle, fat and bone tissue of lambs of the breed German Merino Landsheep in the course of the growing period. J Animal Phys Animal Nutr 90: 385-393, DOI: <u>10.1111/j.1439-0396.2006.00610.x</u>
- Cuthbertson DP (1925) The distribution of phosphorus and fat in the resting and fatigued muscle of the cat, with a note on the partition of phosphorus in the blood. Biochem J 19: 896-910, DOI: <u>10.1042/bj0190896</u>
- Duchesne L, Larson DW (1989) Cellulose and the evolution of plant life. BioSci 39: 238-241, DOI: <u>10.2307/1311160</u>
- Forbes RM, Cooper AR, Mitchell HH (1953) <u>The composition of the adult</u> <u>human body as determined by chemical analysis</u>. J Biol Chem 203: 359-366
- Ghaddar S, Saoud IP (2012) Seasonal changes in phosphorus content of fish tissue as they relate to diets of renal patients. J Renal Nutr 22: 67-71, DOI: <u>10.1053/j.jrn.2011.05.001</u>
- Hennequin M, MCU-PH, Pajot J, MCU, Avignant D, PU (1994) Effects of different pH values of citric acid solutions on the calcium and phosphorus contents of human root dentin. J Endodontics 20: 551-554, DOI: <u>10.1016/S0099-2399(06)80071-3</u>

Handbook on the selection of methods for digestion and determination of total P in environmental samples



- Jacob J, Lawlor DW (1991) Stomatal and mesophyll limitations of photosynthesis in phosphate deficient sunflower, maize and wheat plants. J Exp Bot 42: 1003-1011, DOI: <u>10.1093/jxb/42.8.1003</u>
- Jastrzębska A, Cichosz M, Szłyk E (2010) Simple and rapid determination of phosphorus in meat samples by WD-XRF method. J Analyt Chem 65: 376-381, DOI: <u>10.1134/S1061934810040076</u>
- Kay HD, Byrom FB (1927) <u>Blood-phosphorus in health and disease: I.-</u> <u>The distribution of phosphorus in human blood in health</u>. Br J Exp Pathol 8: 240-253

Koolmann J, Röhm K-H (1998) Taschenatlas der Biochemie. 2. Ed., Thieme-Verlag Stuttgart, 459 pp., ISBN: 3137594022

- Lerch G (1991) Pflanzenökologie. 1. Ed., Akademie-Verlag, Berlin, 535 pp., DOI: <u>10.1002/biuz.19920220413</u>
- Mason HE, Montagna P, Kubista L, Taviani M, McCulloch M, Phillips BL (2011) Phosphate defects and apatite inclusions in coral skeletal aragonite revealed by solid-state NMR spectroscopy. Geochim Cosmochim Acta 75: 7446-7457, DOI: <u>10.1016/j.gca.2011.10.002</u>
- Masuyama R, Nakaya Y, Katsumata S, Kajita Y, Uehara M, Tanaka S, Sakai A, Kato S, Nakamura T, Suzuki K (2003) Dietary calcium and phosphorus ratio regulates bone mineralization and turnover in vitamin D receptor knockout mice by affecting intestinal calcium and phosphorus absorption. J Bone Min Res 18: 1217-1226, DOI: 10.1359/jbmr.2003.18.7.1217
- Mellert K H, Göttlein A (2012) Comparison of new foliar nutrient thresholds derived from van den Burg's literature compilation with established central European references. Eur J Forest Res 131: 1461-1472, DOI: <u>10.1007/s10342-012-0615-8</u>
- Nakamura Y (1982) Effects of dietary phosphorus and calcium contents on the absorption of phosphorus in the digestive tract of carp. Bull Jap Soc Sci Fisheries 48: 409-413, DOI: <u>10.2331/suisan.48.409</u>
- Saunders WMH, Metson AJ (1971) Seasonal variation of phosphorus in soil and pasture. New Zeal J Agricult Res 14: 307-328, DOI: <u>10.1080/00288233.1971.10427097</u>
- Song J-Y, Zhang C-X, Wang L, Song K, Hu S-C, Zhang L (2017) Effect of dietrary calcium levels on growth and tissue mineralization in Japanese seabass, *Lateolabrax japonicus*. Aquacult Nutr 23: 637-648, DOI: <u>10.1111/anu.12431</u>
- Straková E, Suchý P, Karásková K, Jámbor M, Navrátil P (2011) Comparison of nutritional values of pheasant and broiler chicken meats. Ata Vet Brno 80: 373-377, DOI: <u>10.2754/avb201180040373</u>
- Wang Z, Lu J, Yang M, Yang H, Zhang Q (2015) Stoichiometric characteristics of carbon, nitrogen, and phosphorus in leaves of differently aged Lucerne (*Medicaco sativa*) stands. Front Plant Sci 6: 1062, DOI: <u>10.3389/fpls.2015.01062</u>

Handbook on the selection of methods for digestion and determination of total P in environmental samples



- Yoon, G-L, Kim B-T, Kim B-O, Han S-H (2003) Chemical-mechanical characteristics of crushed oyter-shell. Waste Man 23: 825-834, DOI: 10.1016/S0956-053X(02)00159-9
- Zipkin I, McClure FJ, Lee WA (1960) Relation of the fluoride content of human bone to its chemical composition. Arch Oral Biol 2: 190-195, DOI: <u>10.1016/0003-9969(60)90022-4</u>

**For citation:** Baumann K, Zimmer D, Schumann R (*year of download*) Chapter 1.8 Plant and Animal Biomass (Version 1.0) in Zimmer D, Baumann K, Berthold M, Schumann R: Handbook on the Selection of Methods for Digestion and Determination of Total Phosphorus in Environmental Samples. DOI: 10.12754/misc-2020-0001

Handbook on the selection of methods for digestion and determination of total P in environmental samples